

EQUISETIN, AN ANTIBIOTIC FROM
FUSARIUM EQUISETI NRRL 5537,
IDENTIFIED AS A DERIVATIVE OF
N-METHYL-2,4-PYRROLLIDONE

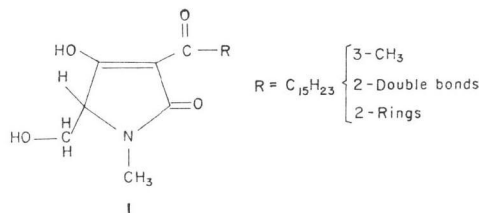
Sir:

Various species of *Fusarium* produce toxins that are implicated in mycotoxicoses in several areas around the world. During a survey of this genus, we found that one species, *Fusarium equiseti* (CORDA) SACCARDO, produced an antibiotic in yields of 5 g/kg when grown on corn grit medium at room temperature (20~24°C). Production, isolation and biological activity of the antibiotic, assigned the trivial name equisetin, have been reported¹⁾. Equisetin is active against several strains of Gram-positive bacteria—*Bacillus subtilis*, *Mycobacterium phlei* and *Staphylococcus aureus*—and the Gram-negative bacteria *Neisseria perflava*, at concentrations of 0.5~4.0 µg/ml of growth substrate; however, it did not inhibit other Gram-negative bacteria tested nor fungi. The LD₅₀ in mice is 63 mg/kg body weight.

Chemical and spectroscopic evidence indicate equisetin to be an N-methyl tetramic acid (1-methyl-3-acyl-5-hydroxymethyl-2,4-dione) as shown in Fig. 1. The proposed structure is unique among metabolites insofar as it contains a hydroxymethyl group at position C₅ which can form an intramolecular hydrogen bond bridge with the acyl carbonyl group of the β-triketone system.

Elemental analysis and high resolution mass

Fig. 1.



spectroscopy of equisetin (m.p. 65~66°C), as well as microanalysis of its copper salt and tetrahydro derivative gave a molecular composition of C₂₂H₃₁NO₄. The UV spectrum of equisetin with characteristic pH-dependent shifts, its IR spectrum (Table 1), formation of its copper salt and positive FeCl₃ and TiCl₃ tests are similar to those given by the acyl tetramic acid, tenuazonic acid²⁾ and the N-methyl tetramic acid, decahydroerythrokyrine³⁾.

Presence of a hydroxymethyl group at C₅ and an N-methyl lactam in the β-triketone structure was indicated by NMR, mass spectroscopy and chemical transformations. The 170.0453 frag-

Fig. 2.

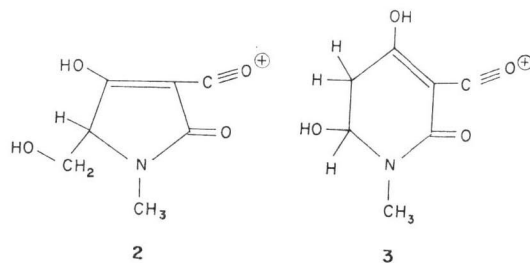


Table 1. Spectral absorption of equisetin, tenuazonic acid, and decahydroerythrokyrine

Compound	UV maxima						IR(CHCl ₃)
	EtOH		Base		Acid		
	λ	εmol	λ	εmol	λ	εmol	
Equisetin ^{a)}	232 292	6,900 10,000	252 292	8,210 7,400	234 294	5,680 8,860	1,680, 1,650, 1,560 cm ⁻¹
Tenuazonic acid ^{b)}	217 277	5,240 13,400	240 279	11,750 14,700	220 277	6,310 12,600	1,735, 1,705, 1,674, 1,630 cm ⁻¹
Decahydroerythrokyrine ^{c)}	225 284	7,250 10,990	246 228	13,800 13,800			1,710, 1,690, 1,635, 1,615 cm ⁻¹

^{a)} Base; 0.01 N KOH-EtOH; acid, 0.05 N H₂SO₄-EtOH.

^{b)} See reference 2.

^{c)} See reference 3.

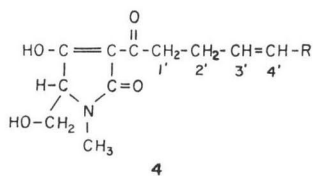
ment in the mass spectrum of equisetin corresponds to $C_7H_8NO_4$. Since this fragment ion contains the nitrogen and all the oxygen of equisetin, it must consist of moiety 2 or 3 as shown in Fig. 2. Reduction with bis-

(2-methoxyethoxy)-aluminum hydride (Red-Al, Aldrich Chemical Co.) results in a cyclic amine as evidenced by the characteristic shift of the NMR N-methyl signal from δ 3.03 to δ 2.43³⁾. The ease of dehydration, which occurs when equisetin is treated with either acetic anhydride in pyridine or *p*-toluenesulfonic acid in boiling benzene, is consistent with a hydroxy group β to a carbonyl function.

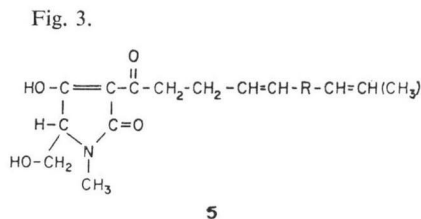
High resolution mass analysis of the dehydrated product gave a molecular ion peak corresponding to $C_{22}H_{29}NO_3$ and a major fragment ion corresponding to $C_7H_8NO_3$. NMR signals at δ 4.58 and δ 4.74 (J_{gem} 2 Hz) are in agreement with a terminal vinyl methylene, which comes from a hydroxymethyl group in moiety 2. The hydroxyl group of moiety 3 would result in a *cis* double bond, which would emit NMR signals similar to those given by the antibiotic mocimycin, a substituted 4-hydroxy-3-propionyl-2(1H)-pyridone⁴⁾. This evidence disproves existence of moiety 3. In addition, the NMR spectrum of equisetin in $CDCl_3$ exhibits an A_2B pattern of the type R_2CH-CH_2R in which the methylene protons appear as a doublet at δ 3.0 and the methine proton as a triplet at δ 3.58 (J 4 Hz). The same protons in deuterated benzene appear as an ABC pattern; (A δ 3.7, B δ 3.06, and J_{AB} 12 Hz, $J_{AC} = J_{BC}$ 4 Hz).

Irradiation of the methine proton at δ 3.06 gave an AB quartet centered at δ 3.6. Molecular models of equisetin indicate that a hydrogen bond could easily be formed between the C_5 hydroxymethyl group and the acyl carbonyl but not the other carbonyls. The hydrogen bond could account for our inability to find an NMR signal attributable to a D_2O exchangeable proton. This is further verified by the consumption of 2 moles of sodium periodate by equisetin as determined by method of AVIGAD⁵⁾.

The mass spectral fragmentation pattern of equisetin indicates the R group of 1 to be $C_{15}H_{23}$. The molecular formula of equisetin implies that the compound must contain, in addition to the



4

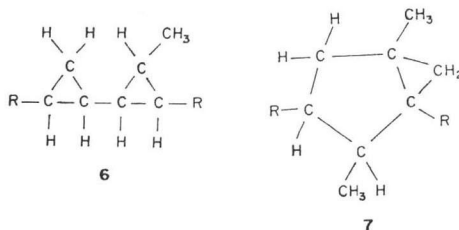


5

β -triketone ring, four double-bond equivalents (DBE). Consumption of 2 moles of hydrogen (product molecular ion $C_{22}H_{35}NO_4$, major fragment in $C_7H_8NO_4$) either over platinum oxide or 5% palladium-on-charcoal in ethanol indicates two reducible functions. One of these, an olefinic moiety of the type $RCCH=CHCH_3$ (δ 1.46, J 4 Hz, C_6D_6) was established by spin decoupling experiments (irradiation of olefinic protons at δ 5.40 collapses the olefinic methyl signals at δ 1.46) and absence of allylic coupling. The other reducible group presumably also involves a double bond because the protons in the vinyl region (δ 5.2~5.4) disappear on hydrogenation. The mass spectra of hydrogenated equisetin showed a series of fragments at m/e 170 ($C_7H_8NO_4$), m/e 185 ($C_8H_{11}NO_4$), m/e 199 ($C_9H_{13}NO_4$) [which were common also to equisetin] and two new fragments at m/e 213 ($C_{10}H_{15}NO_4$), and m/e 227 ($C_{11}H_{17}NO_4$) indicating the formation of two methylene groups. The two fragments at m/e 213 and m/e 227 of hydrogenated equisetin suggests the location of an olefinic bond at C_3' and C_4' as shown in formula 4 (Fig. 3). Also, the fragment at m/e 334 ($C_{19}H_{28}NO_4$) would suggest a loss of a propyl group (C_3H_7) from the fragment m/e 377 ($C_{22}H_{35}NO_4$) which is in agreement for the terminal methyl olefinic moiety ($RCH=CHCH_3$) shown by NMR. Hence, the equisetin structure could be extended to the formula 5 shown in Fig. 3.

The remaining two DBE must then be attributed to ring structures. Methyl signals at δ 0.90,

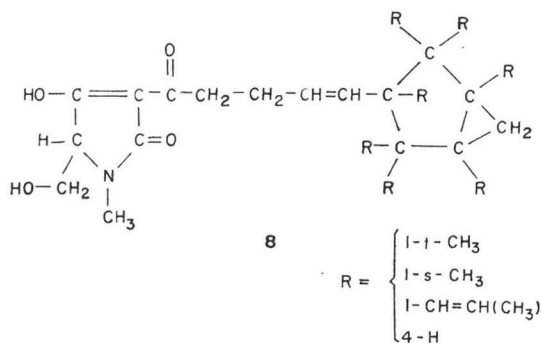
Fig. 4.



6

7

Fig. 5.



6 Hz and δ 1.41 in the NMR spectrum ($CDCl_3$) of equisetin were attributed to secondary and quaternary methyl groups, respectively. KUHN-ROTH analysis gives 1.75 moles of acetic acid accounting for an olefinic and a secondary methyl groups. Values for these groups generally found in the literature are 0.85 and 0.95, respectively. The remaining methyl must be quaternary, which does not react in the KUHN-ROTH procedure. Upon hydrogenation of equisetin in glacial acetic acid at $60^\circ C$, 3 moles of hydrogen were taken up as evidenced by a mass of 379, thus indicating a cyclopropane ring⁶). The NMR of this reduced equisetin product showed two tertiary methyl group signals (δ 1.51, δ 1.54). In view of this evidence, moiety 7 would best support this data because two cyclopropyl rings as shown in moiety 6 (Fig. 4) should yield an additional primary methyl which was not detected in the NMR. This evidence supports the proposed structure 8 (Fig. 5) with the location of the substituents off the fused ring system to be determined.

Periodate oxidation of the sodium salt of equisetin gave a compound (m.p. $111 \sim 113^\circ C$) which chromatographed as a single spot on SiO_2 TLC. It was analyzed as its methyl ester by mass spectroscopy and gave a molecular ion of 419, which corresponds to a formula of $C_{23}H_{33}NO_6$. The mass spectral fragmentation pattern also contained prominent ions, which correspond to $C_{16}H_{24}O$, $C_{15}H_{23}$ and $C_7H_{10}NO_5$. The fragmentation is in accord with the proposed structure 8. The acyltetramic chromophore also has been found in the antibiotics olefin⁷) and

streptolydigin⁸), which are produced by some *Streptomyces* species.

RONALD F. VESONDER
LARRY W. TJARKS
WILLIAM K. ROHWEDDER
HARLAND R. BURMEISTER
JAMES A. LAUGAL*

Northern Regional Research Center,
Agricultural Research, Science and
Education Administration,
U.S. Department of Agriculture,[†]
Peoria, Illinois 61604, U.S.A.
*Eureka College
Eureka, Illinois 61530, U.S.A.

(Received April 19, 1979)

References

- 1) BURMEISTER, H. R.; G. A. BENNETT, R. F. VESONDER & C. W. HESSELTINE: Antibiotic produced by *Fusarium equiseti* NRRL 5537. *Antimicrob. Agents & Chemoth.* 5: 634~639, 1974
- 2) STICKINGS, C. E.: Structure of tenuazonic acid. *Biochem. J.* 72: 332~340, 1959
- 3) SHOJI, J. & S. SHIBATA: The structure of erythrokyrine, a nitrogen-containing colouring matter of *Penicillium islandicum* SOPP. *Chem. Ind.* 1964: 419~421, 1964
- 4) VOS, C. & P. E. J. VERWIEL: The total structure of the novel antibiotic mocimycin (MYC 8003). *Tetrahedron Lett.* 1973: 5173~5176, 1973
- 5) AVIGAD, G.: Rapid, sensitive determination of periodate. *Carbohydr. Res.* 11: 119~123, 1969
- 6) MCCLOSKEY, J. A. & G. H. LAW: Ring location in cyclopropane fatty acid esters by a mass spectrometric method. *Lipids* 2: 225~230, 1966
- 7) GYIMESI, J.; I. OTT, I. HORVÁTH, I. KOCZKA & K. MAGYAR: Antibiotics produced by *Streptomyces*. VIII. A new polyenic antibiotic, olefin, exhibiting antibacterial activity. *J. Antibiotics* 24: 277~282, 1971
- 8) DEBOER, C.; A. DIETZ, W. S. SILVER & G. M. SAVAGE: Streptolydigin, a new antimicrobial antibiotic. I. Biological studies of streptolydigin. *Antibiotics Ann.* 1955/56: 886~892, 1956

[†] The mention of firm names or trade products does not imply that they are endorsed or recommended by the U.S. Department of Agriculture over other firms or similar products not mentioned.